

Potential treatment of Chinese and Western Medicine targeting nsp14 of 2019-nCoV

Chao Liu ¹, Xiaoxiao Zhu ¹, Yiyao Lu ¹, Xu Jia ^{1*}, Tai Yang ^{2*}

¹ Non-coding RNA and Drug Discovery Key Laboratory of Sichuan Province, Chengdu

Medical College, Chengdu, Sichuan, China

² School of Pharmacy, Chengdu Medical College, Chengdu, Sichuan, China

*Correspondence to: Xu Jia: jiaxu@cmc.edu.cn

Tai Yang: taiyang@cmc.edu.cn

Abstract

2019 novel coronavirus (2019-nCoV) caused severe, large-scale acute respiratory disease outbreak in Wuhan, China. The 2019-nCoV has spread to other regions and countries around the world, which is seriously threatening human health. There is an urgent need to develop drugs for the prevention and treatment of 2019-nCoV. 2019-nCoV nonstructural protein 14 (NSP14) carrying RNA cap guanine N7-methyltransferase and 3'-5' exoribonuclease activities could be a potential drug target for intervention. NSP14 of 2019-nCoV shared 98.7% similarity with the one (PDB ID: 5nfy) of acute respiratory syndrome (SARS) Coronavirus. Then, the 2019-nCoV NSP14 structures were modelled by using SARS NSP14 (PDB ID: 5nfy) as template for virtual screening. Based on the docking score, 18 small molecule drugs were selected for further evaluation. The compounds, including Saquinavir, Hypericin, Baicalein and Bromocriptine, could bind the N-terminus and C-terminus of the homology model of the 2019-nCoV Nsp14, thus providing as a candidate drug against 2019-nCoV for further study.

1. Introduction

In December 2019, a large scale, severe acute respiratory disease named as “2019 novel coronavirus (2019-nCoV)” occurred in Wuhan, China, and has already spread to other regions of China and other countries around the world in the following one month, seriously threatening human health. There is an urgent demand to develop drugs for the prevention and treatment of 2019-nCoV. Coronavirus NSP plays an important role in the virus’ genome replication and transcription^{1,2}, and it is generally conserved as an important functional protein in the coronavirus family. Among the family, NSP14 protein has both exonuclease and methyltransferase functions, which is important for replication and transcription of SARS and other coronavirus, thus providing attractive target for drug designs³⁻⁵.

The amino acid sequence alignment revealed that the NSP14 of 2019-nCoV shared 98.7% similarity with the one (PDB ID: 5nfy) of SARS (Figure 1). Thus, the 2019-nCoV NSP14 structures were modelled by using SARS NSP14 (PDB ID: 5nfy) as template. The N-terminus and C-terminus of 2019-nCoV NSP14 were designated as active sites for screening drugs. A total of 7496 drugs obtained in the ZINC database were subjected to the binding screening. Among them, 2100 drugs were approved by FDA, 4264 drugs were approved by other regulatory agencies besides FDA (world-not-FDA) and 1132 drugs are undergoing clinical trials but not yet approved (investigational-only). The docking was carried out using AutoDock Vina1.1.2. Ten top compounds showed the lowest negative vina score in a range of -8.6 to -9.7 kcal/mol were selected from the N-terminal domain of homology model (Table. 1), and eight top compounds with lowest negative vina score in a range of -8.7 to -9.7 kcal/mol were achieved from the C-terminal domain of homology model (Table. 2). More importantly, the compounds, including Saquinavir, Hypericin, Baicalein and Bromocriptine, not only could bind the N-terminus and C-terminus of the homology model (Figure 1. A, B), but also could bind the N- terminal and C-terminal active pockets of the 2019-nCoV Nsp14 (Figure 1. C, D).

2. Materials and Methods

2.1 Docking method

The SARS NSP14 amino acid sequence was downloaded from the PDB protein structure database (PDB ID: 5nfy). The 2019-nCoV amino acid sequence (Accession number: MN908947) was obtained from database of the National Center for Biotechnology Information (NCBI). The homology of above amino acid sequence was aligned using ClustalW. Homology model of the target protein was constructed and optimized by Modeller9.18 using crystal structure of SARS NSP14 (PDB ID: 5nfy) as template. A total of 100 independent structures were constructed, and the one with best DOPE score was selected for further energy minimization by Amber.

The ligands were downloaded from the ZINC database (FDA, world-not-FDA, investigational-only, <http://zinc.docking.org/substances/subsets/>). The 2D structure of the compound was then converted into the corresponding 3D coordinates using the Babel server (<http://openbabel.sf.net>). Then the model was converted to pdbqt format by prepare_receptor4.py script with assigning atom type and partial charge. All rotatable bonds in the ligand were set as flexible for flexible docking. Vina1.1.2 was used for molecular docking. The docking boxes were selected at the N-terminal exonuclease domain (aa: 62-290) and C-terminal transmethylase domain (aa: 291-527) of 2019-nCoV NSP14 respectively.

2.2 Binding free energy calculation

Each simulation system was immersed in a cubic box of TIP3P water with 10 Å distance from the solute. The Na⁺ or Cl⁻ was applied to neutralize the system. General Amber force field (GAFF) 15 and Amber ff14SB force field were used to parameterize the ligand and protein respectively. 10,000 steps of minimization with constraints (10 kcal/mol/Å²) on heavy atoms of complex, including 5,000 steps of steepest descent minimization and 5,000 steps of conjugate gradient minimization, was used to optimize each system. Then each system was heated to 300 K within 0.2 ns followed by 0.1 ns equilibration in NPT ensemble. Finally, 5 ns MD simulation on each system at 300 K

was performed. The minimization, heating and equilibrium are performed with sander program in Amber18. The 5 ns production run was performed with pmemd.cuda. Based on the 5 ns MD simulation trajectory, binding free energy (ΔG) was calculated with MM/GBSA method according to the following equation: $\Delta G_{cal} = \Delta H - T\Delta S = \Delta E_{vdw} + \Delta E_{ele} + \Delta G_{gb} + \Delta G_{np} - T\Delta S$, where ΔE_{ele} and ΔE_{VDW} refer to electrostatic and van der Waals energy terms respectively. ΔG_{gb} and ΔG_{np} refer to polar and non-polar solvation free energies respectively. Conformational entropy ($T\Delta S$) was not calculated for saving time. Besides, the ligands were compared based on the same target, so it is reasonable to ignore the entropy.

3. Results and Discussion

3.1 Docking results of Saquinavir against 2019-nCoV NSP14 model

Saquinavir, as the first FDA-approved HIV protease inhibitor, has been used in the treatment of patients with human immunodeficiency virus (HIV) infection since 1995⁶. Our docking results showed that five of the hydrogen bonds involving ASP-273, ASN-252, ASP-90, and LEU-253 were maintained upon the binding of Saquinavir and N terminus of 2019-nCoV NSP14 (Fig. 1A). Meanwhile hydrogen bonds involving ASN-386, GLN-313, GLY-333 and THR-428 maintained upon the binding of Saquinavir and C terminus of 2019-nCoV NSP14 (Fig. 1C). Saquinavir could bind to the N- and C-terminal active pockets of the 2019-nCoV NSP14 (Fig. 1B, D). The recent study from a drug-target interaction deep learning model showed that Saquinavir can bind to 2019-nCoV RNA-dependent RNA polymerase to inhibit the enzyme activity⁷. Our simulation results showed that Saquinavir can bind two active sites of NSP14, thus Saquinavir could be as a candidate drug against 2019-nCoV for further research.

3.2 Docking results of Hypericin against 2019-nCoV NSP14 model

Hypericin as a main ingredient in traditional Chinese medicine- *Hypericum perforatum* L. (St. John's wort) has been demonstrated activity against RNA viruses in

vitro by inhibiting viral replication⁸. The present docking results show that three of the hydrogen bonds involving ASN-252, GLY-93, and HIS-268 are maintained upon the binding of Hypericin and N-terminus of 2019-nCoV NSP14 (Fig. 2A). The six hydrogen bonds involving ASN-306, ARG-310, ASN-422 and LY-336 are maintained upon the binding of Hypericin and C-terminus of it (Fig. 2C). Hypericin can bind to the N- and C-terminal active pockets of the 2019-nCoV Nsp14 (Fig. 2B, D). Hypericin has been proven to have inhibitory effects on human hepatitis C virus (HCV) and human immunodeficiency virus (HIV)⁹. Combined the present study, Hypericin may have potential antiviral effect against 2019-NCoV. The traditional Chinese medicine-*Hypericum perforatum* L as main composition of Shuanghuanglian oral liquid has been widely used for the treatment of viral influenza. However, Shuanghuanglian oral solution has been suggested for the treatment of 2019-nCoV, triggering a huge crisis of public trust in Chinese scientists. We suggested that anti-2019-nCoV effects of Hypericin should be detected in cell culture models of 2019-nCoV infection. This will help us have a good understanding whether it is a good method to use Chinese medicines for the treatment of 2019-nCoV or not.

3.3 Docking results of Baicalein against 2019-nCoV NSP14 model

Baicalein, a flavonoid compound isolated from the root of *Scutellaria baicalensis* Georgi (Huang Qin in Chinese), inhibit viral replication of parainfluenza, influenza A, hepatitis B, HIV-1, and SARS coronavirus¹⁰⁻¹². The present docking results showed that six of the hydrogen bonds involving ASN-266, ASP-273, GLY-93, GLU-92, and HIS-268 were maintained upon the binding of Hypericin and N-terminus of 2019-nCoV NSP14 (Fig. 2A). Four hydrogen bonds involving ASN-386, ASP-331, and GLN-313 are maintained upon the binding of Hypericin and C-terminus of it (Fig. 2C). Baicalein can also bind to the N- and C-terminal active pockets of the 2019-nCoV NSP14 (Fig. 2B, D). The previous study showed that Baicalein as a novel chemical inhibitor could inhibit ATPase activity of NSP13 protein of SARS coronavirus¹³. The present data suggests that Baicalin may bind to NSP14 protein to exert anti-2019-nCoV activity.

Therefore, we suspect that the anti-2019-nCoV activity induced by the Baicalein could be valuable for further study.

3.4 Docking results of Bromocriptine against 2019-nCoV NSP14 model

Bromocriptine, a specific dopamine receptor agonist for the hypothalamus and pituitary, has inhibitory effect on replication the Dengue virus with low cytotoxicity (half maximal effective concentration, $EC_{50}=0.8-1.6\ \mu\text{M}$; and half maximal cytotoxicity concentration, $CC_{50}=53.6\ \mu\text{M}$)¹⁴. Moreover, Bromocriptine inhibited Zika virus protease activities and exhibited synergistic effects with interferon- $\alpha 2b$ against Zika virus replications¹⁵. It is interesting to find that Bromocriptine can bind to the N- and C-terminal active pockets of the 2019-nCoV NSP14 from our molecular docking results (Fig. 4B, D). The present results showed that three of the hydrogen bonds involving ASN-104, ASP-273 and GLN-145 were maintained upon the binding of hypericin and N terminus of 2019-nCoV NSP14 (Fig. 4A). There was one bond involving THR-428 maintained upon the binding of hypericin and C terminus of it (Fig. 4C).

3.5 The calculation of binding free energy

Based on the 5 ns MD simulation trajectory, binding free energy (ΔG) was calculated by MM/GBSA method. The calculated binding free energies of Saquinavir, Hypericin, Baicalein and Bromocriptine for the N-terminus of the homology model were -37.2711 ± 3.2160 , -30.1746 ± 3.1914 , -23.8953 ± 4.4800 , -34.1350 ± 4.3683 kcal/mol, respectively (Table 3), while the calculated binding free energies were -60.2757 ± 4.7708 , -30.9955 ± 2.9975 , -46.3099 ± 3.5689 , -59.8104 ± 3.5389 respectively, when binding to the C-terminus (Table 4). Taken together, the results demonstrated that Saquinavir had the strong binding free energy.

4. Conclusion

2019-nCoV NSP14, a bifunctional enzyme carrying RNA cap guanine N7-

methyltransferase and 3'-5' exoribonuclease activities could be a potential drug target for intervention. 2019-nCoV NSP 14 shares 98.7% sequence similarity with the corresponding one in SARS. Thus, the homology models of 2019-nCoV NSP14 was structured for virtual screening. Based on the docking score, 18 drugs were selected for further evaluations. Four drugs (Saquinavir, Hypericin, Baicalein and Bromocriptine) could bind to the N-terminal and C-terminal domains of 2019-nCoV NSP 14. Combined the anti-viral function of above four drugs reported by the published literatures, we suggest the anti-2019-nCoV effects of above four drugs should be evaluated in the cell culture models of 2019-nCoV infection.

Acknowledgments

This work was supported by grants from the National Natural Science Foundation of China (NO. 31870135, 31600116) and the “1000 Talent Plan” of Sichuan Province (NO. 980).

References

1. Snijder EJ, Decroly E, Ziebuhr J. The Nonstructural Proteins Directing Coronavirus RNA Synthesis and Processing. *Adv Virus Res* 2016; **96**: 59-126.
2. Sawicki SG, Sawicki DL, Siddell SG. A contemporary view of coronavirus transcription. *J Virol* 2007; **81**(1): 20-9.
3. Chen Y, Tao J, Sun Y, et al. Structure-function analysis of severe acute respiratory syndrome coronavirus RNA cap guanine-N7-methyltransferase. *J Virol* 2013; **87**(11): 6296-305.
4. Eckerle LD, Lu X, Sperry SM, Choi L, Denison MR. High fidelity of murine hepatitis virus replication is decreased in nsp14 exoribonuclease mutants. *J Virol* 2007; **81**(22): 12135-44.
5. Chen Y, Cai H, Pan J, et al. Functional screen reveals SARS coronavirus nonstructural protein nsp14 as a novel cap N7 methyltransferase. *Proc Natl Acad Sci U S A* 2009; **106**(9): 3484-9.
6. Kitchen VS, Skinner C, Ariyoshi K, et al. Safety and activity of saquinavir in HIV infection. *Lancet* 1995; **345**(8955): 952-5.
7. Beck BR, Shin B, Choi Y, Park S, Kang K. Predicting commercially available antiviral drugs that may act on the novel coronavirus (2019-nCoV), Wuhan, China through a drug-target interaction deep learning model. *bioRxiv* 2020.
8. Karioti A, Bilia AR. Hypericins as potential leads for new therapeutics. *Int J Mol Sci* 2010; **11**(2): 562-94.
9. Lenard J, Rabson A, Vanderof R. Photodynamic inactivation of infectivity of human

immunodeficiency virus and other enveloped viruses using hypericin and rose bengal: inhibition of fusion and syncytia formation. *Proc Natl Acad Sci USA* 1993; **90**(1): 158-62.

10. Zhang G-H, Wang Q, Chen J-J, Zhang X-M, Tam S-C, Zheng Y-T. The anti-HIV-1 effect of scutellarin. *Biochem Biophys Res Commun* 2005; **334**(3): 812-6.

11. Sithisarn P, Michaelis M, Schubert-Zsilavecz M, Cinatl J. Differential antiviral and anti-inflammatory mechanisms of the flavonoids biochanin A and baicalein in H5N1 influenza A virus-infected cells. *Antiviral Res* 2013; **97**(1): 41-8.

12. Hour MJ, Huang SH, Chang CY, et al. Baicalein, Ethyl Acetate, and Chloroform Extracts of *Scutellaria baicalensis* Inhibit the Neuraminidase Activity of Pandemic 2009 H1N1 and Seasonal Influenza A Viruses. *Evid Based Complement Alternat Med* 2013; **2013**: 750803.

13. Keum Y-S, Lee JM, Yu M-S, Chin Y-W, Jeong Y-JBotKCS. Inhibition of SARS Coronavirus Helicase by Baicalein. 2013; **34**(11): 3187-8.

14. Kato F, Ishida Y, Oishi S, et al. Novel antiviral activity of bromocriptine against dengue virus replication. *Antiviral Res* 2016; **131**: 141-7.

15. Chan JF, Chik KK, Yuan S, et al. Novel antiviral activity and mechanism of bromocriptine as a Zika virus NS2B-NS3 protease inhibitor. *Antiviral Res* 2017; **141**: 29-37.

Figure and Table legends

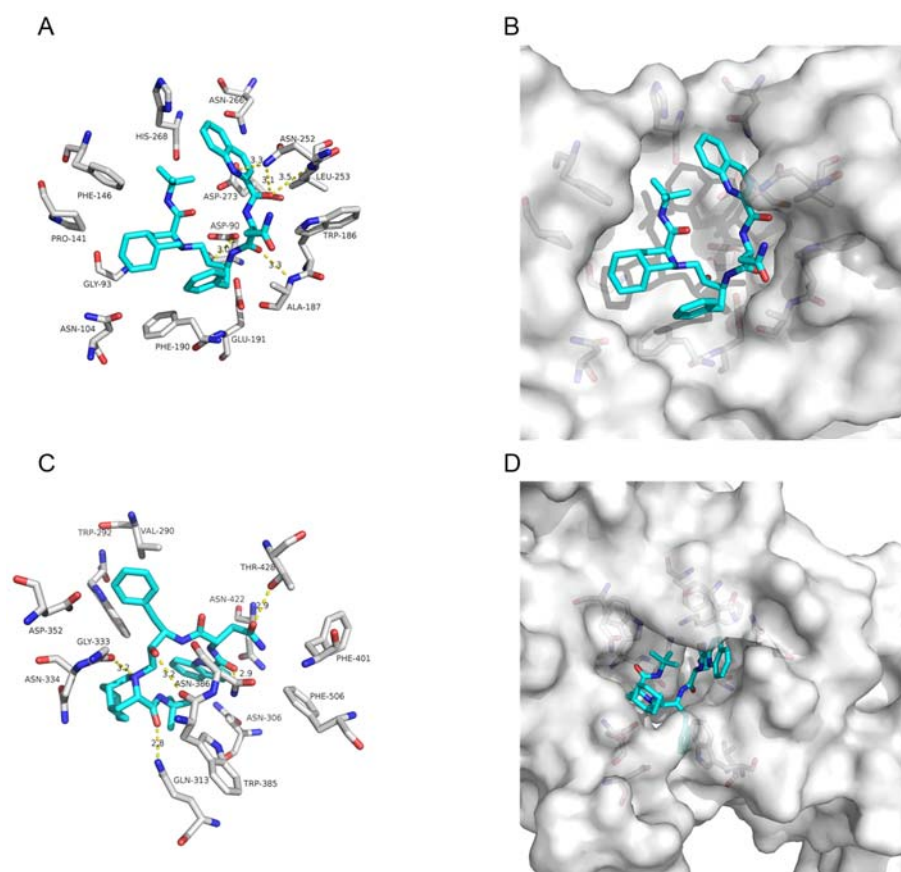


Figure 1. The binding model of Saquinavir against 2019-nCoV NSP14. (A) Interactions between Saquinavir (cyan) and associated residues (off-white) in the N-terminus of the homology model for 2019-nCoV; (B) Binding models of Saquinavir (cyan) in the 2019-nCoV NSP14 protein N-terminus pocket (white surface); (C) Interactions between Saquinavir (cyan) and associated residues (off-white) in the C-terminus of the homology model for 2019-nCoV; (D) Binding models of Saquinavir (cyan) in the 2019-nCoV NSP14 protein C-terminus pocket (white surface). Numbers accompanying dashed yellow lines represents the interaction distance (Å).

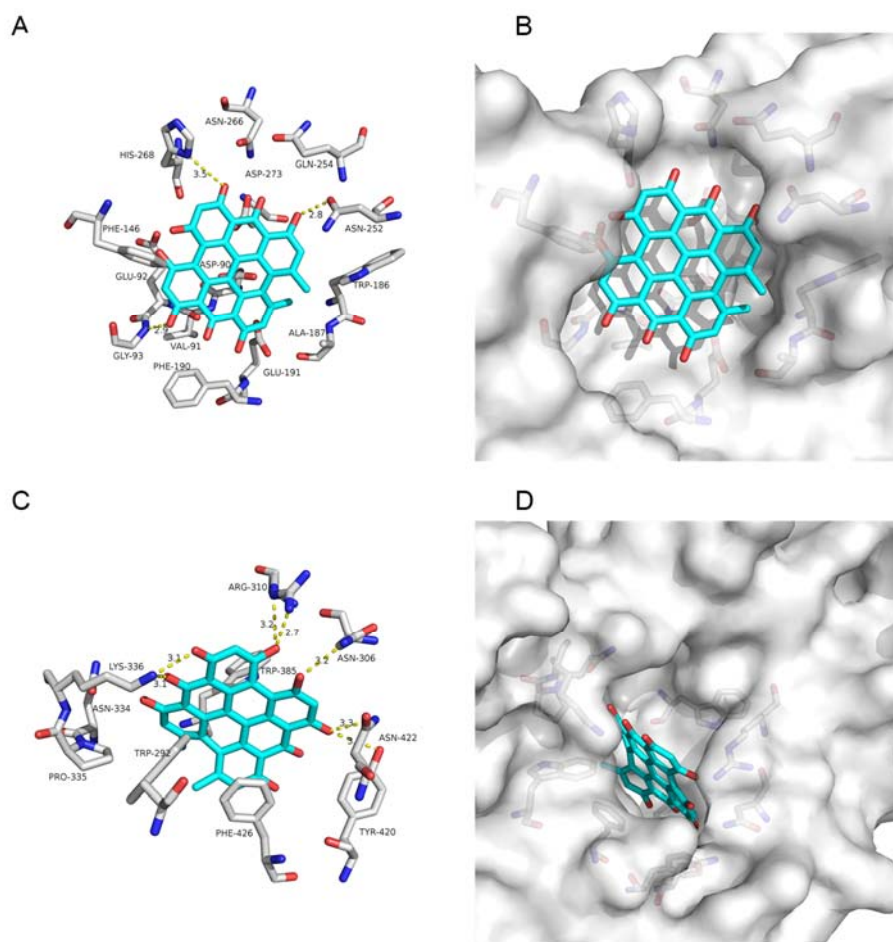


Figure 2. The binding model of Hypericin against 2019-nCoV NSP14. (A) Interactions between Hypericin (cyan) and associated residues (off-white) in the N-terminus of the homology model for 2019-nCoV; (B) Binding models of Hypericin (cyan) in the 2019-nCoV NSP14 protein N-terminus pocket (white surface); (C) Interactions between Hypericin (cyan) and associated residues (off-white) in the C-terminus of the homology model for 2019-nCoV; (D) Binding models of Hypericin (cyan) in the 2019-nCoV NSP14 protein C-terminus pocket (white surface). Numbers accompanying dashed yellow lines represents the interaction distance (Å).

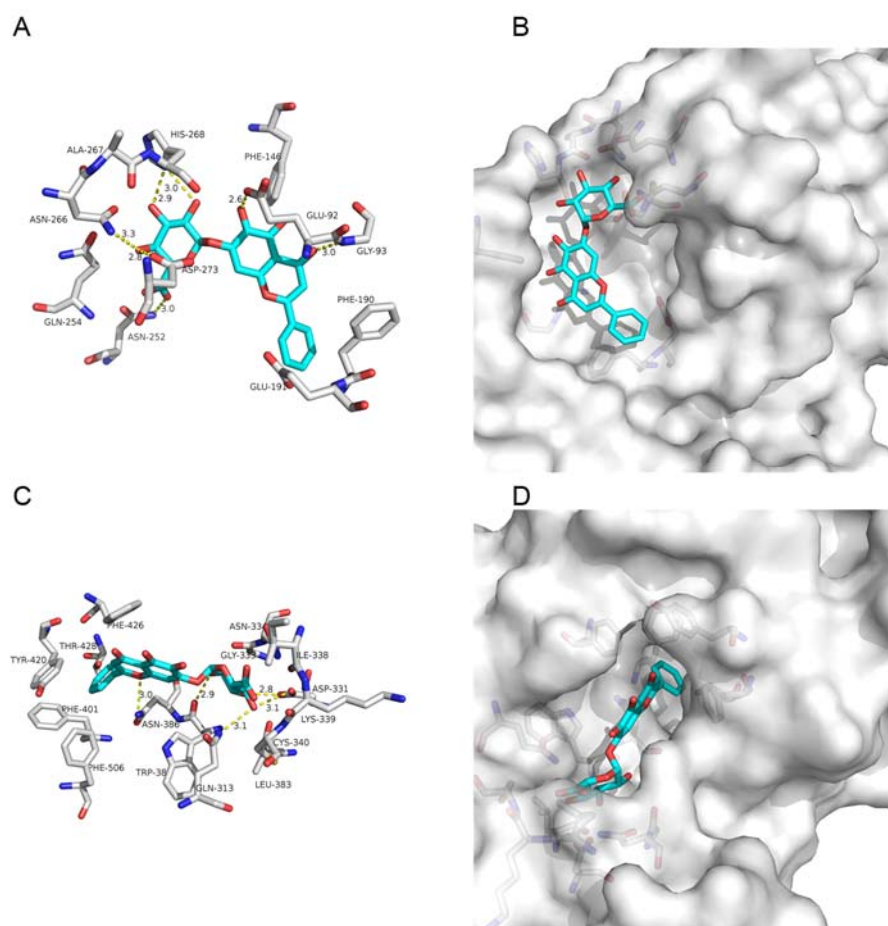


Figure 3. The binding model of Baicalein against 2019-nCoV NSP14. (A) Interactions between Baicalein (cyan) and associated residues (off-white) in the N-terminus of the homology model for 2019-nCoV; (B) Binding models of Baicalein (cyan) in the 2019-nCoV NSP14 protein N-terminus pocket (white surface); (C) Interactions between Baicalein (cyan) and associated residues (off-white) in the C-terminus of the homology model for 2019-nCoV; (D) Binding models of Baicalein (cyan) in the 2019-nCoV NSP14 protein C-terminus pocket (white surface). Numbers accompanying dashed yellow lines represents the interaction distance (Å).

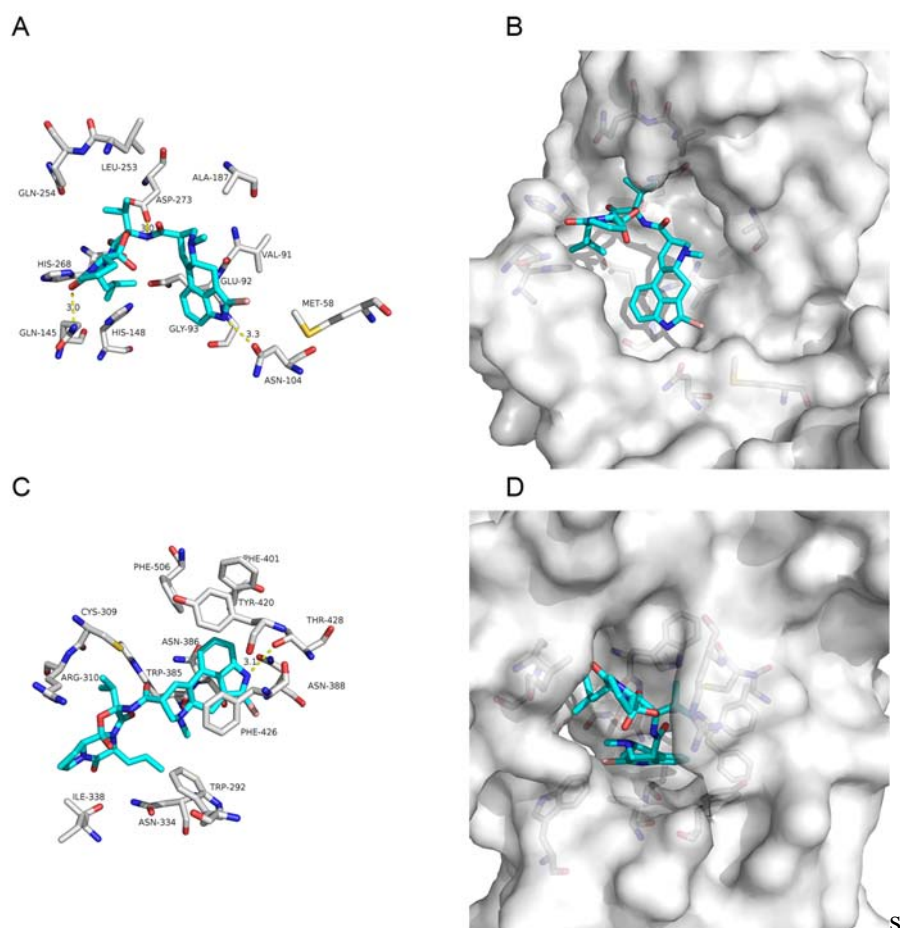


Figure 4. The binding model of Bromocriptine against 2019-nCoV NSP14. (A) Interactions between Bromocriptine (cyan) and associated residues (off-white) in the N-terminus of the homology model for 2019-nCoV; (B) Binding models of Bromocriptine (cyan) in the 2019-nCoV NSP14 protein N-terminus pocket (white surface); (C) Interactions between Bromocriptine (cyan) and associated residues (off-white) in the C-terminus of the homology model for 2019-nCoV; (D) Binding models of Bromocriptine (cyan) in the 2019-nCoV NSP14 protein C-terminus pocket (white surface). Numbers accompanying dashed yellow lines represents the interaction distance (Å).

Table 1 –Ten drugs selected from the N-terminal domain of homology model

Table 1 –Ten drugs selected from the N-terminal domain of homology model			
Drug name	ID	Data	Affinity (kcal/mol)
Hypericin	ZINC000003780340	Investigational-only	-9.7
Bromocriptine	ZINC000053683151	FDA	-9.4
Tanespimycin	ZINC000100014666	Investigational-only	-9.1
Idarubicin	ZINC000003920266	FDA	-9.1
Emend	ZINC000027428713	FDA	-8.9
Baicalein	ZINC000034114798	World-not- FDA	-8.8
Saquinavir	ZINC000029416466	FDA	-8.7
Delavirdine	ZINC000018516586	FDA	-8.7
Silibinin	ZINC000001530850	Investigational-only	-8.6
Golvatinib	ZINC000043195317	Investigational-only	-8.6

Table 2 –Eight drugs selected from the C-terminal domain of homology model

Table 2 –Eight drugs selected from the C-terminal domain of homology model			
Drug name	ID	Data	Affinity (kcal/mol)
Hypericin	ZINC000003780340	Investigational-only	-9.7
Olysio	ZINC000164760756	FDA	-9.4
Sovaprevir	ZINC000085537149	Investigational-only	-9.1
Celsentri	ZINC000003817234	FDA	-9.1
Saquinavir	ZINC000003914596	FDA	-8.9
Maraviroc	ZINC000101160855	World-not-FDA	-8.8
Baicalein	ZINC000034114798	World-not- FDA	-8.7
Bromocriptine	ZINC000053683151	FDA	-8.7

Table 3 –The calculated binding energies of ligand to the N-terminus of 2019-nCoV NSP14

Table 3 –The calculated binding energies of ligand to the N-terminus of 2019-nCoV NSP14				
Energy*	Saquinavir	Hypericin	Baicalein	Bromocriptine
ΔE_{vdw}	-52.9602±2.9999	-36.4737±4.0922	-36.8721±3.4155	-45.8461±3.1764
ΔE_{ele}	-128.6886±21.2732	-78.5578 ±10.7496	-47.8615±12.4900	-119.9028±8.3707
ΔG_{gb}	151.4060±21.5835	90.4227±7.8130	66.0255±11.0420	137.397±6.4239
ΔG_{np}	-7.0283±0.3128	-5.5659±0.2085	-5.1872±0.2001	-5.7839±0.2553
ΔG_{cal}	-37.2711±3.2160	-30.1746±3.1914	-23.8953±4.4800	-34.1350±4.3683

* ΔE_{vdw} = van der Waals energy terms; ΔE_{ele} = electrostatic energy; ΔG_{gb} = polar solvation free energy; ΔG_{np} = nonpolar solvation free energy; ΔG_{cal} = final estimated binding free energy calculated from the above terms (kCal/mol).

Table 4 –The calculated binding energies of ligand to the C-terminus of 2019-nCoV NSP14

Table 4 –The calculated binding energies of ligand to the C-terminus of 2019-nCoV NSP14				
Energy*	Saquinavir	Hypericin	Baicalein	Bromocriptine
ΔE_{vdw}	-70.4383±4.1035	-45.729±2.4822	-48.6473±3.5522	-61.4659±2.9431
ΔE_{ele}	-38.8487±7.5603	-58.4555±12.1238	-192.8463±18.1708	-62.3583±7.0875
ΔG_{gb}	57.7780±6.3018	78.5444±10.3832	202.1598 ±16.7035	70.6739±5.6693
ΔG_{np}	-8.7666±0.3476	-5.3546±0.2317	-6.9761± 0.1614	-6.6602±0.2480
ΔG_{cal}	-60.2757±4.7708	-30.9955±2.9975	-46.3099±3.5689	-59.8104±3.5389